Cover Figure: In our everyday life we encounter oil-water emulsions in many different forms, both in nature (e.g. milk) as well as in industrial products like cosmetics and pharmaceuticals. In order to be stable, oil-water emulsions require some kind of emulsifying agent which prevents the oil phase droplets from coalescing and eventually forming separate oil and water phases. While the liquid fats in milk are stabilized naturally by proteins and protein-like compounds, industrial emulsions usually require high amounts of additives called surface active agents. These surfactants can be hazardous to humans and the environment and it is desirable to substitute them with harmless substances. One approach is to use colloids as emulsifying agents. The front cover shows an image of an oil-water emulsion containing 1% v/v paraffin oil in water with 0.1 % w/w colloidal Ca/AI layered double hydroxide (LDH) as emulsifying agent, taken with the Stony Brook X1A scanning transmission x-ray microscope. Imaging near the calcium-L-absorption edge allows us to map the calcium containing LDH with a spatial resolution of around 70 nm and therefore distinguish LDH from oil and water. When tuning the photon energy to high absorption for calcium (left image), the LDH can be highlighted. For pre-edge energies, the LDH becomes transparent because its absorption coefficient is almost equal to that of water and the oil droplets inside the LDH envelope can be revealed (center image). The right image shows a quantitative calcium map in a range between 0 and 2 micrograms per square centimeter, calculated from the other two images. Studying these emulsions in their natural hydrated state at atmospheric pressure with high spatial resolution is important to better understand the stabilization mechanism and its dependencies and compare it to existing models from colloid chemistry. To our knowledge, the information obtained is currently not accessible by any other techniques.

U. Neuhaeusler (SUNY Stony Brook, U. Goettingen), S. Abend (U. Kiel), C. Jacobsen (SUNY Stony Brook), G. Lagaly (U. Kiel)

This work was supported by a fellowship for PhD studies from German Academic Exchange Service (U.N.), by a travel grant from Boehringer Ingelheim fund for basic medical research (S.A.) and by the Office of Biological and Environmental Research, U.S. DOE under contract DE-FG02-89ER60858.

## **DISCLAIMER**

This report was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government nor any agency thereof, nor any of their employees, nor any of their contractors, subcontractors, or their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commerical product, process, or service by trade name, trademark, manufacturer, or otherwise, does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency, contractor, or subcontractor thereof. The views and opinions of authors express herein do not necessarily state or reflect those of the United States Government or any agency, contractor, or subcontractor thereof.

Printed in the United States of America Available from National Technical Information Service U.S. Department of Commerce 5285 Port Royal Road Springfield, VA 22161